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ABSTRACT

Abstract: The present investigation was designed to assess the protective capabilities of Ficus carica (fig) extract and olive oil, administered individually and in combination, as potent natural antiproliferative agents against the adverse effects of DMBA on the HBP.

Material and methods: Fifty five-week-old male Syrian hamsters weighing between 80 and 120 grams were split up into five groups of ten hamsters within every group: GI: left untreated, GII: 7, 12-dimethyl benz[a]anthracene (DMBA) applied topically only on three occasions per week during 14 weeks at a concentration of 0.5% in liquid paraffin, GIII: DMBA (0.5% in liquid paraffin) applied topically on three occasions per week during 14-week plus oral fig latex (50 g lyophilized fig latex in 50 ml distilled water at a dosage of 1000 mg/ml shortly prior to experimental usage). GIV: DMBA (0.5% in liquid paraffin) used topically on three occasions per week over 14-week plus oral EVOO alone (7.6 ml/kg b.w by oral

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gavage, before and/or after DMBA application. GV: DMBA applied topically (0.5% in liquid paraffin, three occasions per week during 14 weeks) alongside oral fig latex and EVOO administration using dosages and administration techniques comparable to those used in individual therapies. The Qur'anic proportion of one fig to seven olives served as the basis for the specific dosages for each fig extract and olive oil.

Results: The current research findings demonstrated that fig latex and EVOO had a positive regression impact on the growth of tumors. The combination of fig latex and EVOO significantly decreased the occurrence of OSCC (oral squamous cell carcinoma) from 100% to 0%. According to the immunohistochemical study, fig latex and EVOO significantly reduced the expression of the antiproliferative marker PCNA in comparison to GII. Such findings highlighted the chemopreventive potential of fig latex and EVOO, suggesting their roles in inhibiting tumor growth and inducing apoptosis in oral cancer cells.

Conclusion: This study underscores the value of natural dietary components in cancer prevention and supports the further exploration of fig latex and EVOO as novel, safe, and effective therapeutic agents for oral malignancies..

Keywords: : HBP carcinoma, fig latex, EVOO

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1. INTRODUCTION

Oral squamous cell carcinoma (OSCC) represents significant global public health concern because of its high prevalence worldwide. According to GLOBOCAN 2020 estimates, approximately 377,713 cases of OSCC are newly diagnosed each year, and 177,757 deaths occur annually worldwide ⁽¹⁾. In Egypt, Abd El-Aziz et al. (2020)⁽²⁾ study among 1664 smoker participants revealed that 1.5% had OSCC and 0.12% had verrucous carcinoma.

Oral carcinogenesis is a multifactorial disease arising from the interaction of lifestyle, environmental, genetic, and epigenetic factors⁽³⁾. 7,12-Dimethylbenz(a)anthracene (DMBA)-induced oral carcinogenesis in golden Syrian hamsters which serves as an established widely laboratory paradigm for examining molecular, biochemical, histological, and immunohistochemical (IHC) changes ⁽⁴⁾. DMBA is commonly utilized to induce tumor development in the golden Syrian hamster buccal pouch (HBP)⁽⁵⁾. HBP, a pocket-like anatomical structure, develops a well-established OSCC following repeated topical DMBA applications. DMBA initiates neoplasia by causing significant inflammation and dysplasia within the buccal pouches, alongside extensive oxidative damage to DNA ⁽⁶⁾. Cumulative data indicate that DMBA-induced oral tumors share histological, morphological, biochemical, and molecular characteristics with human oral tumors ⁽⁷⁾.

There is considerable interest in identifying potential protective agents capable of mitigating the adverse effects linked to toxins and chemotherapy ^(8, 9). Consequently, medicinal plants are utilized for treating various ailments and counteracting the side effects of chemotherapeutic agents, including cardiovascular and renal complications ^(10, 11).

According to the Holy Qura'n (Surah 95. At-Teen) "By the fig and the olive, By mount Sinai, By this city of security⁽¹²⁾. The olive is referenced seven times in the holy Quran—six times expressly and one time implicitly—while the fig is mentioned just once ⁽¹³⁾.

Within the herbal remedies that are regularly researched and shown to have advantageous anti-inflammatory, immunomodulatory, antibacterial, anticancer, chemopreventive, analgesic, and antioxidant benefits are olive (Olea Europe) and fig (Ficus carica). Oleic acid and phenolic substances are primarily responsible for their biological characteristics (14-

Fig (Ficus carica) and olive (Olea europaea) are regarded as inexpensive raw materials that serve as valuable sources for high added-value products, particularly phenolics (17). Fig and olive leaves are readily accessible, low-cost natural resources that potentially contain a comparable abundance of health-promoting bioactive phytochemicals (18).

A mixture of figs and olives exhibited enhanced antioxidant stability and bioavailability compared to individual plant extracts. This is likely due to the greater variety of polyphenols in the mixed extracts, which can interrupt detrimental oxidative reactions by donating a hydrogen atom ^(19, 20).

There is no published report on the in vivo chemopreventive influence of consuming olive oil and fig extract separately or in combination as strong natural antiproliferative resources towards the harmful side effects of DMBA on the HBP

2. MATERIAL AND METHODS:

Chemicals: DMBA (0.5%) was purchased dissolved in paraffin oil from Sigma Aldrich. Extra Virgin Olive Oil (EVOO), certified organic (NOP/EU), with a density of 0.910 g/ml (Product No: W530191), was also purchased from Sigma Aldrich

Company (Saint Louis, MO 63103, USA).

Preparation of fig latex: Ficus carica latex was harvested by collecting drops following the incision of young fig tree leaves at a private farm in New Damietta City, Damietta, Egypt. The fresh latex was subsequently filtered using a $0.22\mu m$ filter and preserved at -20° C. For the experiment, the latex was prepared as a suspension of 50 g fig latex in 50 ml distilled water, yielding a concentration of 1000 mg/ml, immediately prior to its administration.

Animals: Fifty male Syrian hamsters five-week-old weighing between 80 and 120 grams, were taken from the animal department at Cairo University (Cairo, Egypt). The hamsters were kept in typical Sawdust-bedded enclosures having regulated humidity (30–40%), warmth ($20\pm2^{\circ}$ C), and a 12-hour light/dark cycle. All hamsters were provided with conventional feed and water ad libitum. A healthy animal exhibited a regular, smooth walk, clear, bright eyes, skin that is healthy, and a silky, glossy coat without swellings, injuries, parasites, and dry areas. There are no cuts or scratches on their mouths.

Sample size: According to the Abd Al-Wahab et al. (2020) investigation ⁽²¹⁾, a sample size of 10 animals per group was determined for the present investigation. This size provides 80% power to identify a 0.53 average variance at a confidence interval of 95% and a two-tailed significance level (alpha) of 0.05. In 80% of the experiments (the power), the resulting p-value was below 0.05 (two-tailed), classifying the results as "statistically significant." Conversely, in the remaining 20% of the experiments, the mean difference was considered "not statistically significant." The report was generated using GraphPad StatMate 2.00.

Experimental design: Following a one-week acclimatization period, the hamsters were randomly assigned to five groups (G(s)) of ten hamsters within every group: **GI (negative control)** left without therapy; GII (DMBA alone) 0.5% DMBA in paraffin oil applied topically to the right HBP utilizing a size 4 camel hair brush, on three occasions weekly during 14 weeks ⁽²¹⁾. **GIII (fig latex)** DMBA applied topically (as GII) plus oral gavage of fig latex suspension (1000 mg/ml concentration of 50 g lyophilized fig latex in 50 ml distilled water) immediately before experimental use, administered before and/or after DMBA application ⁽²²⁾. **GIV (EVOO):** Topical application of DMBA (as GII) + Oral administration of EVOO alone (7.6 ml/kg b.w by oral gavage, before and/or after DMBA application) ⁽²²⁾. **GV (fig latex - EVOO)** received topical DMBA (as GII) plus oral gavage of fig latex and EVOO, using the same doses and administration protocol as the individual treatments. The chosen doses of fig extract and olive oil were based on the Qur'anic ratio of one fig to seven olives

General health assessments: Throughout the trial, changeover in the animals' overall health status were documented. Indicators of illness or injury in animals included one or more of the following symptoms: anorexia, lethargy, corner huddling, sneezing, wheezing, nasal or ocular discharge, perianal wetness, diarrhea, and alopecia. Animal body weights were recorded weekly during the experimental period.

Investigations: Upon completion of the trial, gross findings (including size of the mucosa, exudation, ulceration, and tumors) were documented. The hamsters were then euthanized, the right cheek pouch was removed, and the size of every tumor was calculated using a Vernier caliper. Tumor volume (Vmm³) was determined using the formula Vmm³ = $(4/3) \pi$ [(D1/2) (D2/2) (D3/2)], where D1, D2, and D3 represent the three diameters (in mm) of the tumor ⁽²³⁾. Subsequently, the right cheek pouch was removed, fixed in 10% neutral buffered formalin, and treated and inserted into paraffin blocks on a regular basis for subsequent histological and immunohistochemical analysis.

Histopathological examinations: The preserved samples were incorporated in paraffin wax to create paraffin blocks after being dehydrated in an increasing ethanol sequence. A rotary microtome was utilized to generate 4μ m-thick tissue slices, which were then placed on glass slides, processed, and stained using hematoxylin and eosin (H&E) to light microscopic analysis.

Calculation of the Depth of Invasion (DOI): Depending on the H&E slide, DOI was calculated for every surgical sample. DOI was calculated from the lowest point of tumor penetration to the basal layer of the surface epithelium. The American Joint Committee on Cancer (AJCC) has additionally classed it as less invasive (≤5 mm), moderately invasive (6–10 mm), and highly invasive (≥10 mm) (24) (Fig.1). The Leica QWIN V3 computer program for photo analysis (Switzerland), was used to calculate the DOI. This was carried out in the Department of Oral and Dental Pathology, Faculty of Dental Medicine (Boys-Cairo), Al-Azhar University, Egypt.

Immunohistochemical examination: To show the expression of PCNA antibodies, additional tissue slices were sliced and subjected to the typical marked streptavidin-biotin procedure. The tissue slices fixed in paraffin were dewaxed and rehydrated using distilled water and graded ethanol. After ten minutes of treatment with 3% H2O2 in methanol, endogenous peroxidase was inhibited. After applying citrate buffer solution (pH 6.0) and microwaving for three periods of five minutes at 95°C, the antigen was recovered. Phosphate-buffered saline (PBS) was then used for rinsing. Next, single or double drops of the main antibody (PCNA) diluted 1:100 in Tris buffer solution were added to the tissue sections. They spent the entire night at normal temperature at 4°C in a humid environment. Following a PBS wash, a biotinylated secondary antibody was incorporated and allowed to sit at an ambient temperature for 30 minutes. Tissue slices were rinsed with PBS and then exposed to diaminobenzidine (Sigma, USA) for two to four minutes in order to produce color. Once the

desired color strength had been attained, the slides were cleaned, Hematoxylin was used as a counterstain, and after that a mounting media was applied.

To ascertain the occurrence of positive instances and the exact spots of immunostaining within the tissues. A light microscope was used to examine the immune-stained slices. Additionally, the area percentage of immunostaining-positive cells was calculated using a picture analytics computer program.

Statistical analysis: The findings were statistically assessed and documented as the mean \pm standard deviation (SD). SPSS version 17.0 for Windows was utilized to perform a one-way test of variation (ANOVA). ANOVA was utilized to compare greater than two independent groups utilizing quantitative data and parametric distribution. The LSD test was then used for post hoc analysis. P > 0.05 was deemed non-significant, p < 0.05 was deemed significant, and p < 0.001 was deemed extremely significant.

3. RESULTS:

Gross observations and tumor volume: GI (negative group) Animals showed no gross alterations, maintaining healthy and active behavior; the length of both buccal pouches was approximately 5 cm. The right HBP mucosa was pink, surface was smoothness, and don't have notable abnormalities (Fig.2A). GII (DMBA-treated group) Animals exuded a foul odor and white particles from their mouths. All hamsters had noticeable perioral hair loss, which extended up to the belly in some animals with substantial body weight. This might have been brought on by inadequate food intake as a result of DMBA-caused oral cavity inflammation. The pouch depth started to drop by up to 2 cm and didn't change till the trial was over. The pouch depth began to decrease up to 2 cm and remained fixed until the end of the experiment. The animals were debilitated and skinny. The right HBP mucosa showed a whitish membrane and granular surface on the pouch mucosa, erythema of various degrees, and numerous elevated nodules also present. Using vernier caliper, hamsters with tumors had an average tumor volume of 775.5 mm³ (Fig.2B). In GIII (fig latex treated group) and GIV (EVOO treated group), HBP mucosa showed variable changes, including different exophytic tumor mass, erythematous mucosal surface to usual regular color, and the hamsters seemed healthy. Using vernier caliper, the average tumor volume in hamsters with tumors in GIII and GIV were 93.8 mm³ and 90.2 mm³ respectively (Fig.2C&D). GV (combination group) The hamsters seemed healthy, and the erythematous mucosal surface of the HBP mucosa changed to a standard typical hue having a smooth surface. (Fig.2E). General health examinations and common clinical findings observed within the present research groups are summarized in Table 1.

Table 1: Typical clinical results seen in the groups under study.

features	Groups							
icatures	GI	GII	GIII	GIV	GV			
loss of appetite	-	+++	+	+	-			
Inactivity	-	+++	+	+	-			
corner huddling	-	+++	+	+	-			
sneezing, wheezing, and/or discharge from the nose or eyes,	-	+++	+	+	-			
wetness around the tail, diarrhea	-	+++	+	+	-			
hair loss	-	+++	+	+	-			
Gross observations								
Papillomatous lesion	-	+++	+	+	-			
Pouch length	5 cm	1.5-2cm	4cm	4cm	4.5cm			

Ulcers	-	+++	+	+	-
Exudation	-	+++	+	+	-
Tumors	-	100 %	30 %	30 %	-

⁻⁼ no change; += mild; ++= moderate; +++= severe

Statistical analysis results, with regard to the tumor volume, revealed that GII recorded greatly significant variation with GIII, GIV and GV (p < 0.001). Furthermore, GIII, GIV and GV displayed non-significant difference when comparing to each other (p value > 0.05). (**Table 2, Fig.1**).

Statistical analysis results, with regard to body weight, revealed that, there was greatly significant increase in GI, GIII, GIV and GV when compared with GII (p < 0.001). Moreover, no significant variations were shown amongst other groups ($p \times 0.05$). (**Table 2, Fig.1**).

GI GII GIII **GIV** GV F P-value (n = 10)(n = 10)(n = 10)(n = 10)(n = 10)Mean 775.5 93.8 90.2 **Tumor** 0.001 134.8 volume HS ±SD 109.3 47.1 40.5 495 296 412 420 487 Mean **Body** 0.001 10.11 weight HS 13.3 22 $\pm SD$ 18 16.1 **17**

Table.2: Comparison across examined groups as regard tumor volume and Body weight.

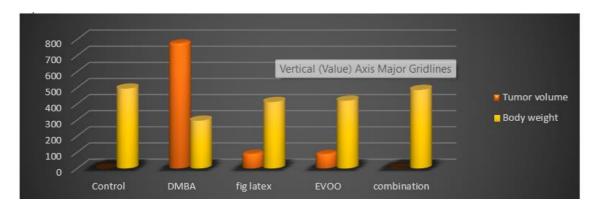


Fig.1: Bar chart representing tumor volume and body weight in the studied groups

Histopathologic findings and immunohistochemical (IHC) results: GI The HBP mucosa's lining epithelium was extremely thin, having keratinized stratified squamous epithelium that is three to five cells thick, according to H&E staining. One to two layers of polyhedral spinous cells, one layer of cuboidal basal cells, and flattened granular cells with small keratohyalin granules were visible in the epithelial layers. The epithelium was sparsely coated with keratin. The epithelium connective tissue interaction was reasonably flat without rete processes. The submucosa was composed of delicate and loose connective tissue and a layer of striated muscle fibers (**Fig. 2F**). The IHC staining in **GI** showed that although a small number of PCNA-positive cells were discovered and dispersed across the basal cell layer, PCNA antigen was only isolated in the nuclei of epithelial cells. The cytoplasm showed no response. PCNA exhibited positive nuclear expression (10.14%) throughout the basal cell layer (**Fig. 3A**).

In **GII**, the H&E stain indicated that two instances had mild SCC, and eight cases had well-differentiated SCC. Histologically, well-differentiated SCC shows malignant epithelial cells that penetrate extensively through the connective tissue (DOI=11.3mm). The invading epithelial cells were observed as cell nests or as detached and scattered cells, with confirmation of keratinization. The nuclear/cytoplasmic ratio was demonstrated in tumor cells having pleomorphic,

hyperchromatic nuclei. (**Fig.2G**). In **GII**, the IHC staining utilizing PCNA showed positive nuclear expression (72.53%) across the layers of the epithelium (**Fig.3B**).

In **GIII** and **GIV**, the H&E stain revealed exhibited hyperkeratosis to mild epithelial dysplasia in two hamsters. While six animals had moderate to severe epithelial dysplasia alongside top-to-bottom alterations or carcinoma in situ (CIS), whereas two animals had superficial invasion of malignant cells in a type of well-differentiated SCC that was juxtapithelial and was not spreading to the deeper C.T. (DOI=0.9-1.1mm). (**Fig.2H&I**). In **GIII** and **GIV**, the IHC staining via PCNA showed positive nuclear expression (33.5% and 30.22% respectively) across the layers of the epithelium (**Fig.3C&D**).

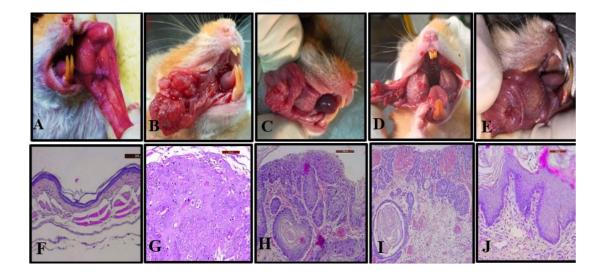
In **GV** four animals displayed mild to moderate epithelial dysplasia, characterized by focal dysplastic areas within the epithelium, including hyperchromatism, altered nuclear/cytoplasmic (N/C) ratio, cellular and nuclear pleomorphism, prominent nucleoli, and multiple foci of group cell keratinization. The remaining six animals showed hyperkeratosis with normal epithelium. (**Fig.2J**). In **GV**, the IHC staining utilizing PCNA showed positive nuclear expression (21.41%) across the layers of the epithelium (**Fig.3E**). Histopathological alterations in the HBP of the control and treated animals among the studied groups are summarized in Table 3.

Table 3: Histopathological alterations observed in the HBP of the control and treated animals across the study groups.

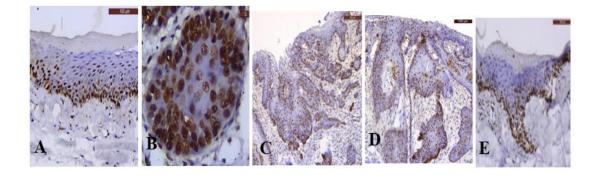
Features	Groups						
reatures	GI	GII	GIII	GIV	GV		
Irregular epithelial stratification	-	+++	+	+	+		
Basal cell hyperplasia	-	+++	+	+	-		
loss of basal cells polarity	-	+++	+	+	-		
Individual cell keratinization	-	+++	+	+	+		
keratin pearls	-	+++	+	+	-		
Nuclear pleomorphism	-	+++	+	+	+		
Cellular pleomorphism	-	+++	+	+	+		
Abnormal mitotic figures	-	+++	+	+	+		
Hyperchromatism	-	+++	+	+	+		
Abnormal nucleus/cytoplasm ratio	-	+++	+	+	+		
Inflammatory infiltration	-	+++	+	+	-		
Increase amount of collagen fibers	-	-	+	+			
DOI	-	11.3mm	0.9mm	1.1mm	-		
Epithelial histological changes							
Mild dysplasia	-	+++	20%	20%	40%		
Moderate dysplasia	-	+++	40%	30%	-		

Severe dysplasia	-	+++	10%	20%	-
CIS	-	+++	20%	10%	-
SCC	-	Well (70 %) Moderate (50 %)	Well (20 %)	Well (20 %)	

⁻⁼ no.change; += mild; ++= moderate; +++= severe.



(Fig.2A): Photograph of GI (normal). Displaying a rosy, smooth buccal pouch mucosa. (Fig.2B): Photograph of GII (DMBA) several polypoid papillary tumor masses enveloped by hemorrhage spots. (Fig.2C): Photograph of GIII (fig latex group) mucosa showed varied shifts, including smooth, normal-colored, reddened mucosal surface. (Fig.2D): Photograph of GIV (EVOO group) displaying a little lump that is neither bleeding or ulcerating. (Fig.2E): Photograph of GV (combination group) displaying a significant reduction in tumor mass size without bleeding or ulceration. (Fig.2F): Photomicrograph of GI (normal) displaying: The epithelium is composed of 2-4 layers: 5 superficial keratinized squamous cells with flattened rete ridges, the C.T layer, the muscular layer, and the deep layer of loose areolar connective tissue. (H&E stain X200) (Fig.2G): Photomicrograph of GII (DMBA) displaying moderately-differentiated SCC with deeply invasive tumor islands into the connective tissue. (H&E stain X200). (Fig.2H): Photomicrograph of GIII (fig latex group) displaying well-differentiated SCC having superficial invasion of tumor islands into the connective tissue. (H&E stain X200). (Fig.2J): Photomicrograph of GV (combination group) showing mild to moderate epithelial dysplasia. (H&E stain X200).



(Fig.3A): IHC expression of PCNA in GI shows positive nuclear expression in basal epithelial layers. (Fig.3B): IHC expression of PCNA in GII displaying positive nuclear expression across the tumor cells. (Fig.3C): IHC expression of

PCNA in GIII displaying positive nuclear expression across the layers of the epithelium. (**Fig.3D**): IHC expression of PCNA in GIV displaying positive nuclear expression across the layers of the epithelium. (**Fig.3E**): IHC expression of PCNA in GV displaying positive nuclear expression across the layers of the epithelium.

Statistical analysis findings of PCNA expression and DOI:

According to the findings of statistical analysis, GI had the smallest mean area percentages (10.14%) and GII had the greatest mean area percentages (72.53%) for PCNA expression. (**Table 4, Figure 4**).

The DMBA treated group (GII) when compared with either GI (left untreated) or chemopreventive groups (GIII, GIV and GV), there was highly significant variation; (p value< 0.001). Furthermore, GIII, GIV and GV showed non-significant difference when comparing to each other (p value > 0.05). (Table 5).

The findings of the statistical analysis showed that GII had the greatest mean area percentages (11.3%) and GIII had the smallest mean area percentages (0.9%) in terms of DOI expression (**Table 4, Figure 4**).

There was a highly significant variation (p value<0.001) between the DMBA-treated group (GII) and either the GI (left untreated) or chemopreventive groups (GIII, GIV, and GV). Furthermore, GIII, GIV and GV showed non-significant difference when comparing to each other (p value > 0.05). (**Table 5**).

Tuble 112 of the Chiptesbion comparisons across the groups under study.									
		GI	GII	GIII	GIV	GV		P-	
		(n = 10)	F	value					
PCNA	Mea n	10.14	72.53	33.5	30.22	21.41	11.0	< 0.001 HS	
	±SD	2.16	12.54	16.1	12.2	8.7			
DOI	Mea n	-	11.3	0.9	1.1	0	180. 7	< 0.001	
	±SD	-	2.5	0.88	1.07	0	/	HS	

Table 4: DOI and PCNA expression comparisons across the groups under study.

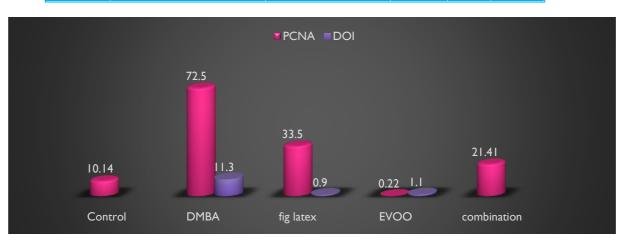


Fig.4: Bar chart representing PCNA experssion and DOI in the studied groups.

P-value for post analysis using LSD PCNA GIV GV Mean ± SD Range G1 GII GIII 7.2 - 13.20.000 10.14 ± 2.16 0.000 0.000 0.000 Group I Group II 72.53 ± 12.54 71.3 - 79.30.0000.0000.000 0.000 Group III 33.5 ± 16.1 28.9 - 37.30.0000.0000.125 0.117 25.4 - 34.2 $30..22 \pm 12.2$ 0.000 0.111Group IV 0.0000.12516.9 - 27.3Group V 21.41 ± 8.7 0.000 0.000 0.1170.11111.02 P-value <0.001 (HS)

Table 5: Comparing the groups under study in terms of PCNA

4. DISCUSSION:

Animal models are thought to be crucial for studying the course of illnesses and the development of diagnostic or treatment procedures. There has been increased interest in the utilization of herbal remedies for the chemoprevention and treatment of oral cancer, including green tea and fruit and vegetable preparations from grape, apple, and tomato (25-27). To present, the anticancer efficacy of fig latex and EVOO, a therapeutic natural plant, on oral cancer hadn't been documented. The chemopreventive properties of fig latex and EVOO were investigated in this work utilizing an animal model that was developed for mouth cancer utilizing DMBA.

Combining therapy of two or three polyphenols reduced the development of cancer more successfully than a single chemical therapy, according to preclinical research on the anticancer impact of polyphenols from various plants. numerous metabolic pathways implicated in carcinogenesis might be pleiotropically affected by the synergistic impact of numerous nutrients, which can also influence the bioavailability of natural substances ^(28, 29). Thus, using an animal model of oral cancer, we assessed the impact of fig latex and EVOO on cancer cells. The current research demonstrated the inhibitory impact of fig latex and EVOO on the growth of malignant tumors by comparing average tumor size and IHC staining using PCNA between the fig latex-treated, EVOO- treated either alone or combined and cancerous group. Based on available English literature, this represents the first investigation to evaluate fig latex and EVOO as a novel chemopreventive strategy for DMBA-induced HBP carcinogenesis.

According to reports, the OSCC occurrence in the HBP was highly replicable (100%) after 14 weeks of DMBA (0.5%) treatment in hamsters ⁽²⁶⁾. DMBA can cause various dysplastic and neoplastic lesions having molecular and morphological changes which can be connected to human carcinogenesis, according to the dosage and time of the carcinogen being delivered ⁽³⁰⁾. After 14 weeks of cancer initiation, DMBA caused precancerous and cancerous morphological alterations on the HBP within the present investigation.

DMBA treatment to induce cancer in animals may additionally contribute to various clinical findings including loss of appetite, Inactivity, coughing, sneezing, and/or nasal or ocular discharge, hair loss as well as substantial body weight loss, not being able to eat, an absence of hunger, and an elevated rate of metabolism due to the occurrence of oral cancer (25, 31). The use of DMBA in this investigation resulted in a notable decrease in body weight and great changes in the animal's overall health of the cancer-induced animals, which is consistent with other results. However, the fig latex and/or EVOO therapy regulated the weight loss.

Ghandehari et al $(2018)^{(32)}$ stated that, cancer cells have been shown to be able to avoid apoptosis and to fail at any stage of the cell cycle. Therefore, it is probable that proliferation of cells and tumor diameters are decreasing because to the activation of apoptosis with fig latex and EVOO with cell cycle. In the current study, fig latex and EVOO decreased the tumor voulmes of fig latex and EVOO treated animals either alone or combined with highly significant reduction (p < 0.001) in comparison to the tumor sizes in the malignant group of hamsters.

Our results revealed that fig latex and EVOO were able to downregulate PCNA expression either alone or combined with significant variation in comparison to GII (p value < 0.001). In accordance, fig latex and EVOO have been researched as a possible antioxidant and anti-inflammatory medications due to their polyphenol compositions which trigger apoptosis and prevent cell division (33, 34). Numerous plants showed antitoxic action for various types of cancer, such as the colon, ovary, liver, kidney, central nervous system, and stomach cells exposed to various toxic chemicals, according to reporters

experiments ^(35, 36). These researchers attributed this effect to the antiproliferative properties of the plants. Additionally, the synergies between fig latex and EVOO may contribute to their antitumor effectiveness ^(11, 22, 33, 34). Furthermore, hydroxytyrosol, one of the strongest antioxidants in olive oil, and oleuropein, the main phenol in olive goods, could further work by altering oncogenic signaling pathways, which results in cell death ⁽³⁷⁾. Furthermore, only a small number of investigations show that the various constituents of fig latex might inhibit and reduce inflammation and tumor development ^(32, 38, 39). AlGhalban et al (2021)⁽³⁹⁾ stated that, fig latex exhibits diverse molecular mechanisms of action, such as antiproliferative and antimetastatic effects, as well as notable impacts on cellular morphology. Additionally, Ghandehari et al (2018)⁽³²⁾ found that, fig latex protects against the damaging effects of oxidative stress brought on by free radicals generated by immune system cells.

According to Tezcan et al (2015)⁽⁴⁰⁾ and Tezcan et al (2019)⁽⁴¹⁾, the expression of lethal-7d (let-7d) is induced by Fig latex and oleuropein (olive oil extract), which in turn prevents invasion. Additionally, it was found that the use of fig latex and olive oil extract have an inhibitory effect on the growth of endothelial cells cultivated in a collagen matrix-containing three-dimensional medium.

Fig latex and EVOO have been found to possess the ability to impede the growth of cancerous cell lines through various mechanisms⁽⁴²⁻⁴⁴⁾. Notably, this effect was observed without any accompanying cytotoxicity towards normal human cells. Fig latex and EVOO have been found to possess antiproliferative effects on cancer cells, specifically its ability to induce apoptosis through modulation of the cell cycle. This is achieved by increasing the population of cells in the G0/G1 phase while decreasing the population in the S and G2/M phases^(32, 45). According to Han et al (2023)⁽⁴⁵⁾ and Shin et al (2017)⁽⁴⁶⁾, another possible mechanism for these effects could be connected to the inhibition of DNA synthesis, the beginning of apoptosis, the arrest of the cell cycle in cancerous cells, and activation of both the extrinsic death receptor and the intrinsic mitochondrial-dependent apoptosis signaling pathway.

Unexpectedly, blending fig with EVOO offers a novel cure that is enriched in polyphenolic compounds, capable of contributing to a hydrogen atom and preventing the adverse reaction activities. In addition to attenuating cardiac and renal toxicity, this argued therapy additionally demonstrated strong anti-inflammatory, antioxidant, and anti-apoptotic properties. As a result, it gained considerable interest for usage in avoiding some common chronic illnesses, such as diabetes and hepatic diseases (11, 19, 47).

5. CONCLUSIONS

Based on the current findings, we conclude that fig (Ficus carica) tree latex and EVOO are promising candidates as potential agents for inhibiting the proliferation and development of cancerous cells, and may demonstrate comparable effectiveness in clinical investigations

REFERENCES

- [1] Sung H, Ferlay J, Siegel R, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2021;71(3):209-49.
- [2] El-Aziz A, AbouShousha A, Ali S, Zahran F. Prevalence of potentially malignant lesions and oral cancer among smokers in an Egyptian cohort: a hospital-based cross-sectional study. Adv Dent J. 2020;2(3):93-100.
- [3] Mello F, Melo G, Pasetto J, Silva C, Warnakulasuriya S, Rivero E. The synergistic effect of tobacco and alcohol consumption on oral squamous cell carcinoma: a systematic review and meta-analysis. Clin Oral Investig. 2019;23(7):2849-59.
- [4] Manoharan S, Vasantha S, Silvan S, Baskaran N, Singh A, Kumar V. Carnosic acid: a potent chemopreventive agent against oral carcinogenesis. Chem Biol Interact. 2010;188(3):616-22.
- [5] Karthikeyan S, Srinivasan R, Wani S, Manoharan S. Chemopreventive potential of chrysin in 7,12-dimethylbenz(a)anthracene-induced hamster buccal pouch carcinogenesis. Int J Nutr Pharmacol Neurol Dis. 2013;3(1):46-53.
- [6] Manoharan S, Karthikeyan S, Essa M, Manimaran A, Selvasundram R. An overview of oral carcinogenesis. Int J Nutr Pharmacol Neurol Dis. 2016;6(2):51-62.
- [7] Tanaka T, Ishigamori R. Understanding carcinogenesis for fighting oral cancer. J Oncol. 2011;2011:1-10.
- [8] Rashid S, Ali N, Nafees S, Ahmad S, Hasan S, Sultana S. Abrogation of 5-fluorouracil-induced renal toxicity by bee propolis via targeting oxidative stress and inflammation in Wistar rats. J Pharm Res. 2013;7(2):189-94.
- [9] Osama A, Engy R, Gehad E. Immunomodulatory effect of artichoke (Cynara scolymus) on carbon tetrachloride-induced immunosuppression in rats. Ann Vet Anim Sci. 2014;1:66-76.
- [10] Khalil S, Mohammed A, Abd El-fattah A, Zaglool A. Intermediate filament protein expression pattern and

- inflammatory response changes in kidneys of rats receiving doxorubicin chemotherapy and quercetin. Toxicol Lett. 2018;288:89-98.
- [11] Elghareeb M, Elshopakey G, Hendam B, Rezk S, Lashen S. Synergistic effects of Ficus Carica extract and extra virgin olive oil against oxidative injury, cytokine liberation, and inflammation mediated by 5-Fluorouracil in cardiac and renal tissues of male albino rats. Res Pharm. 2021;28:4558-72.
- [12] Sheikh D, Dixit A. Plants in the holy Quran: a look. World J Pharm Pharm Sci. 2015;4(8):715-38.
- [13] Marwat S, Khan M, Khan M, Fazal-ur-Rehman F, Akbari A, Mushtaq A, et al. Medicinal and pharmacological potentiality of the plant At-Tîn-common fig (Ficus carica L.). Asian J Chem. 2011;23(1):1-10.
- [14] Vallejo F, Marín J, Tomás-Barberán F. Phenolic compound content of fresh and dried figs (Ficus carica L.). J Food Comp Anal. 2012;130(3):485-92.
- [15] Erol Ö, Arda N. Phenols of virgin olive oil protects nuclear DNA against oxidative damage in HeLa cells. Food Chem Toxicol. 2012;50(10):3475-9.
- [16] Bahadori S, Salamzadeh J, Kamalinejad M, Ardekani M, Keshavarz M, Ahmadzadeh A. Study of the effect of an oral formulation of fig and olive on rheumatoid arthritis (RA) remission indicators: a randomized clinical trial. Iran J Pharm Res. 2016;15(3):537.
- [17] Şahin S, Bilgin M. Olive tree (Olea europaea L.) leaf as a waste by-product of table olive and olive oil industry: a review. J Sci Food Agric. 2018;98(4):1271-9.
- [18] Yousef N, El-Ghandour A, El-Shershaby S. Antimicrobial activity of fig and olive leaves extracts. J Food Sci. 2019;10(12):503-8.
- [19] Bashandy M, El-Rasheid A, Hasan H, Fathya A. Protective and therapeutic effects of olive oil and Ficus carica as natural antioxidants on some biochemical parameters in liver of γ-irradiated male albino rats. Arab J Biol Sci. 2014;25(1-C):1-16.
- [20] Rubió L, Macià A, Castell-Auví A, Pinent M, Blay M, Ardévol A, et al. Effect of the co-occurring olive oil and thyme extracts on the phenolic bioaccesibility and bioavailability assessed by in vitro digestion and cell models. Food Chem. 2014;149:277-84.
- [21] Al-Wahab A, Gobran H, Dameer H. Effect of rosmarinic acid as a chemopreventive modality on experimentally induced hamster buccal pouch carcinogenesis. J Evid Based Complement Altern Med. 2020;66(4):2411-21.
- [22] Gafar A, Khalifa M, Elshaer F. Protective and therapeutic effects of olive oil and Ficus carica on the histological and cytological injuries of liver and kidney in γ-irradiated male albino rats. Arab J Biol Sci. 2018;29(2-C):69-88.
- [23] Silvan S, Manoharan S. Apigenin prevents deregulation in the expression pattern of cell-proliferative, apoptotic, inflammatory and angiogenic markers during 7,12-dimethylbenz[a]anthracene-induced hamster buccal pouch carcinogenesis. Arch Oral Biol. 2013;58(1):94-101.
- [24] Faisal M, Abu Bakar M, Sarwar A, Adeel M, Batool F, Malik K, et al. Depth of invasion (DOI) as a predictor of cervical nodal metastasis and local recurrence in early stage squamous cell carcinoma of oral tongue (ESSCOT). PLoS One. 2018;13(8):e0202632.
- [25] Elagouz M, Zahran A, Zouair M, Abd-Alhafez A. Therapeutic potential of epigallocatechin-3-gallate on cancer stem cells of DMBA induced hamster buccal pouch carcinoma. Arab J Dent Sci. 2023;26(4):517-30.
- [26] Alqalshy E, Ibrahim A, Abdel-Hafiz A, Kamal K, Alazzazi M, Omar M, et al. Effect of docosahexaenoic acid as a chemopreventive agent on experimentally induced hamster buccal pouch carcinogenesis. Food Sci. 2022;31:100558.
- [27] Shamia A, Abd-Alhafez A, Zouair M. Chemopreventive and therapeutic efficacy of thymoquinone on experimentally induced hamster buccal pouch carcinogenesis. Int J Health Sci. 2022;6(S7):2716-34.
- [28] Niedzwiecki A, Roomi M, Kalinovsky T, Rath M. Anticancer efficacy of polyphenols and their combinations. Nutr J. 2016;8(9):552.
- [29] Papakonstantinou A, Koumarianou P, Diamantakos P, Melliou E, Magiatis P, Boleti H. A systematic ex-vivo study of the anti-proliferative/cytotoxic bioactivity of major olive secoiridoids' double combinations and of total olive oil phenolic extracts on multiple cell-culture based cancer models highlights synergistic effects. Nutr J. 2023;15(11):2538.
- [30] Mohammed E, Ahmed M, Zouair M. Synergistic effect between cetuximab and curcumin on experimentally induced hamster buccal pouch carcinoma. Arab J Dent Sci. 2017;20(2):145-52.
- [31] Zhang W, Yin G, Dai J, Sun Y, Hoffman R, Yang Z, Fan Y. Chemoprevention by quercetin of oral squamous

- cell carcinoma by suppression of the NF- κB signaling pathway in DMBA-treated hamsters. Anticancer Res. 2017;37(8):4041-9.
- [32] Ghandehari F, Fatemi M. The effect of Ficus carica latex on 7,12-dimethylbenz(a)anthracene-induced breast cancer in rats. J Appl Pharm. 2018;8(4):286.
- [33] Ibrahim H, Rabeh N, Elghandour H, Sabry S. The functional role of dried fig, date and olive oil on rats with immune dysfunction. J Food Sci. 2018;9(9):313-20.
- [34] Fathy A, Naji R, Bashandy M. Antioxidant and hepatoprotective effects of fig fruit extract with olive oil and date-palm fruit extract on hepatic toxicity of oral subchronic exposure to some nanoparticles in Wistar rats. J Food Qual. 2023;2023(1):7584688.
- [35] Govindan S, Jayabal A, Shanmugam J, Ramani P. Antioxidant and hepatoprotective effects of Hypsizygus ulmarius polysaccharide on alcoholic liver injury in rats. J Food Sci Wellness. 2021;10(4):523-35.
- [36] Fathy S, Mahmoud M. Moringa oleifera Lam. leaf extract mitigates carbon tetrachloride-mediated hepatic inflammation and apoptosis via targeting oxidative stress and toll-like receptor 4/nuclear factor kappa B pathway in mice. J Food Sci Wellness. 2021;10(3):383-91.
- [37] Casaburi I, Puoci F, Chimento A, Sirianni R, Ruggiero C, Avena P, et al. Potential of olive oil phenols as chemopreventive and therapeutic agents against cancer: a review of in vitro studies. Nutr J. 2013;57(1):71-83.
- [38] Almehmadi A. Biological activities of Ficus carica L. fig leaves and latex. J Food Sci. 2023;14(7):173-9.
- [39] AlGhalban F, Khan A, Khattak M. Comparative anticancer activities of Ficus carica and Ficus salicifolia latex in MDA-MB-231 cells. Saudi J Biol Sci. 2021;28(6):3225-34.
- [40] Tezcan G, Tunca B, Bekar A, Yalcin M, Sahin S, Budak F, et al. Ficus carica latex prevents invasion through induction of let-7d expression in GBM cell lines. J Funct Foods. 2015; 35:175-87.
- [41] Tezcan G, Aksoy S, Tunca B, Bekar A, Mutlu M, Cecener G, et al. Oleuropein modulates glioblastoma miRNA pattern different from Olea europaea leaf extract. J Funct Foods. 2019;38(9):1102-10.
- [42] Hashemi S, Abediankenari S, Ghasemi M, Azadbakht M, Yousefzadeh Y, Dehpour A. The effect of fig tree latex (Ficus carica) on stomach cancer line. Iran Red Crescent Med J. 2011;13(4):272.
- [43] Aysin F, Özek N, Acet N, Koc K. The anti-proliferative effects of Ficus carica latex on cancer and normal cells. J Appl Environ Sci. 2015;9(2):145-51.
- [44] Kahraman G, Özkaya M, Yıldırım Ö. Potential anti-cancer effects of extra virgin olive oil and its phenolic extracts on hepatocellular carcinoma cells. Int J Nutr Sci. 2023;7(2):112-22.
- [45] Han S, Hou H, Zhang Y, Fang Z, Li Y, Zhang L, et al. Oleuropein and its hydrolysate from extra virgin olive oil inhibit breast cancer cells proliferation interfering with the PI3K-AKT signal pathway. Food Chem. 2023; 110:105880.
- [46] Shin B, Lee S, Moon S, Han S, Hwang E, Kim S, et al. Latex of Ficus carica L. induces apoptosis through caspase and Bcl-2 family in FaDu human hypopharynx squamous carcinoma cells. Food Chem. 2017;42(4):183-90.
- [47] Fathy A, Bashandy M, Bashandy S, Mansour A, Azab K. The beneficial effect of natural antioxidants from olive oil with fig and date palm fruit extracts on biochemical and hematological parameters in rats treated with doxorubicin and γ-radiation. Food J. 2018;3(1):722-35.