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Short Communication

Does GATA3 act in tissue-specific pathways? A meta-analysis-based approach

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Abstract

The GATA3 transcription factor is expressed in many tissues such as the immune system, kidney, brain, endometrium, and mammary epithelial cells. As such it must co-ordinate a diverse transcriptional program to achieve specific outcomes in different tissues. One of the most interesting questions raised is whether GATA3 will be involved in the same pathways in every tissue or will be involved in distinct regulatory networks within different tissue types? While previous studies may imply the latter, with some known targets of GATA3 perhaps being specific to cell-type or tissue-type, the question has not been systematically addressed until now. With the advent of techniques such as co-expression meta-analysis a better understanding of the pathway partners of GATA3 can be obtained and specifically the partners within different tissue types can be found, yielding leads for future studies. Here, a recent technique of meta-analysis from the Oncomine database has been employed to probe this very question. Data obtained implies that GATA3 is involved in distinct pathways in different tissue types.

Keywords: GATA3, meta-analysis, tissue-specific, transcription, co-expression, p63

Introduction

The GATA3 transcription factor has been studied intensively in the immune system, but has most recently been shown to be important in the context of breast cancer and the estrogen receptor α pathway.^[1,2] These and many other studies have shown the power of linking breast cancer co-expressed gene datasets of GATA3 to its ability to play a putative role in the luminal A subtype of breast cancers, while also having a physiological role in normal breast epithelial cells. Recently, it has been proposed that GATA3 might act differently in different tissues.^[3] As meta-analysis of GATA3 co-expressed genes in breast cancers, from the Oncomine database, yielded data that was analogous to previously known studies, it was possible that this technique could also be utilized to show tissue-specific (or common) pathways of GATA3.^[4]

Consistent co-expression of genes has been shown to link their

protein products to the same pathways, although it is impossible to determine the nature of this association (e.g., upstream, downstream, together in a complex) without further study. What this does generate, however, are rich leads into previously unknown or undefined areas for study.

The Oncomine meta-analysis technique^[4–6] was employed using various studies of different types (leukemia, lymphoma, brain, kidney, and prostate studies), all of which have been previously observed to show GATA3 expression. Co-expressed genes of GATA3 were then assessed within these cancer/tissue-types and are described below. Finally, the gene lists generated for individual analyses were compared to assess common co-expressed genes over multiple tissues. Tentative results indicate that GATA3 may be involved in distinct pathways in different tissue-types, confirming the initial hypothesis. A prime example of this is the estrogen receptor alpha (ER α) pathway in breast cancer studies,^[4] where ER α is the highest co-expressing gene

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with GATA3, whereas in the other cancer types investigated, this high level of co-expression was not evident, thereby indicating that the ER α pathway need not play a significant role for GATA3 in every situation.

Co-expression Meta-analysis of Leukemia Studies

GATA3 expression has previously been observed in leukemia cells, however, the role GATA3 may play has not been intensively investigated, leaving a need for new knowledge.^[7]The highest co-expressing gene with GATA3 in leukemia meta-analysis was the T-cell receptor (TCR) alpha locus ($TRA(\hat{a})$), in over 60% of studies [Additional File 1]. This is confirmation of a previous report that demonstrated GATA3 binding to the TCR-alpha enhancer.^[8] Similar to this, the TCRdelta locus (TRD@) was also co-expressed with GATA3 and has also previously shown to be bound by GATA3.^[9,10] Both of these results act as initial validation of the meta-analysis technique. As might be expected, there was an abundance of immunological co-expressed genes such as these. Interleukins and their receptors (IL32, IL27RA, IL2RB, IL10RA, IL7R and IL15), other T-cell receptors and associated factors (TRBC1, TRAT1, LAT, CD3D, CD3E, CD3G, TARP, ICOS, MAL, TRGV9, TRGC2 and PTCRA), and granzyme proteases (GZMA, GZMB, GZMK and GZMM) are just some notable examples of the many immune genes represented.

As GATA3 has been intensively investigated in an immunological context for many years, especially with regard to T-cells, these data may help to shed light on the role GATA3 plays, albeit by implication via these co-expressed genes. For example, the granzyme proteases have been shown to be involved in cell-death pathways, regulated by cytotoxic T-cells and natural killer cells.^[11]

Co-expression Meta-analysis of Lymphoma Studies

As lymphomas are also derived from the cells of the immune system, it is no surprise that the co-expressed genes with GATA3 have a significant overlap with those of the leukemias. For example, the interleukins and receptors (*IL15*, *IL15RA*, *IL7R*, *IL2RB*, *IL32*, *IL1R1*, *IL3RA*, *IL1R2*, *IL18R1*, *IL13RA1* and *IL10*) show overlap with 4 of the 6 co-expressed for leukemias [Additional File 2].

Chemokines and receptors are also notably co-expressed with GATA3 in lymphomas (*CXCL10*, *CXCR6*, *CXCL9*, *CXCL11*, *CXCL12* and *CXCL13*) as are the members of the STAT transcription family (*STAT1*, *STAT3*, *STAT4*, *STAT5A* and *STAT5B*). It is of interest that GATA3 has previously been observed to co-operate with STAT5A in naïve CD4 T-cells to prime towards a Th2 phenotype, emphasizing its role in differentiation.^[12]

Co-expression Meta-analysis of Brain Studies

GATA3 has been seen to be important in the development of the mammalian brain, with *Gata3* -/- mice displaying severe brain and nervous system abnormalities.^[13] GATA3 was also shown to be downstream of GATA2.^[14,15] More recently, however, it was shown that the role of GATA3 in neonatal (i.e. post-development) mouse brain is limited.^[16]

It is possible that in conditions such as brain tumors the cancer will be in a de-differentiated state, thus expressing more developmental proteins such as GATA3. The highest co-expressing gene with GATA3 in Oncomine brain studies was the Disco Interacting Protein 2, homolog C (*DIP2C*) [Additional File 3]. While this remains little-studied, it is interesting to note that dip2 is expressed in the mouse nervous-system.^[17] Of interest, the neurofibromin 2 (*NF2*) gene is strongly expressed in the developing brain^[18] and is involved in schwannomas^[19] and is here also co-expressed with GATA3 in Oncomine brain studies. Whether GATA3 plays a role in schwannomas remains to be investigated.

Co-expression Meta-analysis of Kidney Studies

The hypoparathyroidism, sensoneural deafness, and renal abnormality (HDR) syndrome in humans is linked to haploinsufficiency of GATA3.^[20–22] GATA3 is also expressed in developing human and mouse kidneys.^[23,24] Specifically, GATA3 was shown to be important for the development of the nephric duct.^[25]These data show a requirement for GATA3 to be expressed within renal tissues.

For meta-analysis, however, there were a limited number (9) of Oncomine renal studies with co-expressing genes for GATA3. Even so, many highly co-expressing genes were observed such as KCNJ1 (ROMK), which is seen with GATA3 in over half of these studies [Additional File 4]. ROMK, as might be expected by its function as a potassium channel, is important to kidney function.^[26,27] Furthermore, GATA3 co-expressed with many other transporters (SLC7A8, SLC19A2, SLC12A1, SLC7A7, SLC27A2, SLC26A7, SLC29A2, SLC25A33, SCN3A, SCN2A, KCNK10, KCNJ15, KCNJ10, CLIC5, CLCNKA, CLCNKB, CACNG4, ATP1A1, ATP6VOA4 and AQP2), possibly implying a GATA3 pathway in the kidney. There was also a modest co-expression with renin (REN), which as part of the renin-angiotensin system is one of the key genes of the kidney.

Co-expression Meta-analysis of Prostate Studies

The role of GATA3 in the prostate is the least studied of all tissues presented. In contrast, however, there were many (17) Oncomine studies with co-expressed genes for GATA3 [Additional File 5]. In a recent study of candidate prostate cancer genes, GATA3 was identified and confirmed by immunohistochemistry.^[28] This report showed that GATA3 expression is normally low in normal prostate, perhaps explaining why it has not yet been studied to any significant degree. However, as Oncomine studies are based on cancer, the data presented here might be most useful in analysis of GATA3 in prostate cancer. GATA sites were also shown to be important for full induction of prostate-specific antigen (PSA) in LNCaP cells,^[29] while a different antigen, the prostate stem cell antigen (PSCA) is co-expressed with GATA3 in the meta-analysis.

As GATA3 is co-expressed with keratin 18 in breast cancers,^[4] it is of interest that in prostate, GATA3 co-expressed with many other keratins (*KRT5*, *KRT7*, *KRT13*, *KRT14*, *KRT15*, *KRT17*, and *KRT23*). GATA3 was also co-expressed with transcription factors including p63 (*TP63*). The p63 protein is essential for the development of stratified epithelia in tissues such as the prostate and is highly expressed in proliferative epithelial stem cells.^[30] GATA3 and p63 appeared to be overexpressed in urothelial cancers in a recent study combining microarrays and histochemistry, although this was not observed in prostate.^[31] Other co-expressed transcription factors from the current meta-analysis include *SMARCD3*, *SMAD3*, *PDLIM3*, *PDLIM4*, *NR4A2*, *NR4A3*, *MEIS1*, *ZNF593*, *ZNF516*, *ZNF423*, *ZEB1*, *TEAP2A*, *TCF21*, *KLF6*, *KLF9*, *JUN*, *JUNB*, *ESR2*, *FHL1*, and *FHL2*.

Conversely to p63 the ZEB1 factor is involved in epithelial to mesenchymal transition in carcinogenesis, i.e., the dedifferentiation of epithelial cells^[32] and is a putative prostate cancer biomarker.^[33] It remains to be investigated how GATA3 might interplay in prostate transcriptional networks with the aforementioned co-expressed factors. However, it is of interest that ZEB1 protein can bind *in-vitro* to a GATA3 gene silencer and can repress GATA3 in reporter gene assays, in the context of the JurkatT-cell line.^[34]While this needs to be shown in a more rigorous manner in an *in-vivo* context using, for example, chromatin immunoprecipitation of ZEB1 on the GATA3 regulatory element, whether this can also be shown in prostate is a point of significant interest.

pS2 (*TFF1*) is a co-expressed gene of GATA3 in breast cancers.^[4] Interestingly, this was again seen to be the case in prostate cancers. The presence of prostate pS2 has previously

been investigated, correlating with prostate hyperplasia and cancer.^[35,36] Although it remains to be shown, it is possible that pS2 may then be a common target of GATA3 in both breast and prostate tissues.

However, the most frequent co-expressing gene with GATA3 was *TRIM29* (ATDC) in almost 60% of studies analyzed. ATDC was observed to be downregulated in cancers including prostate cancers,^[37] and when transfected into cancer cells, ATDC reduced colony formation in soft agar,^[38] which is suggestive of a tumor suppressive role.

Altogether, the genes listed in this meta-analysis may prove to be most useful in future studies of GATA3 in prostate, acting as a "road-map" of putative pathway-partners. For example, GATA3 might act in co-operation with p63 in epithelial cells, as GATA3 itself has been shown to be important for epithelial terminal differentiation, albeit in the context of mammary glands.^[39,40]

Common GATA3 Co-expressing Genes

All of the previous meta-analyses were compared for common overlapping genes [Table 1]. As GATA3 co-expressed genes in breast cancer, using an identical technique, have already been reported,^[4] the overlaps with these new meta-analyses are shown here in [Additional File 6].

As can be observed, only 10 genes were co-expressed with GATA3 in over 2 tissues (*TRA@*, *PDE4DIP*, *MAL*, *LCK*, *KRT7*, *HLA-DQB1*, *IL32*, *IL15*, *DPYD*, and *CH13L2*). This low number reflects the concept of tissue-specific pathways of GATA3 regulation, supported by the types of genes co-expressed with GATA3 within the tissues themselves (e.g., immunological genes in leukemias and lymphomas, transporters in the kidney studies). Altogether, the data not only provide evidence that GATA3 may be involved in distinct pathways, but also reveal some of these pathway partners.

Conclusions

While the *in-silico* meta-analysis data presented can do no more than imply pathway partners of GATA3 in various tissues, the data presented can be considered as initial leads into future analysis. While an in-depth meta-analysis yields much data, it can only scratch the surface regarding putative pathways of GATA3. These data also strongly imply tissue-specific functions of GATA3, consistent with mounting evidence from the literature. How GATA3 can act in such a tissue-specific manner is an issue that has not yet been addressed, but likely to involve unique tissue-specific partners and post-translational modifications, controlling the functions of this transcription factor.

Table 1: Common co-expressed genes ofGATA3 over different tissues

	Leukemia	Lymphoma	Kidney	Prostate	Brain		Leukemia	Lymphoma	Kidney	Prostate	Brain
TRA@	\checkmark	✓			\checkmark	ILIRLI			~		✓
PDE4DIP	\checkmark	\checkmark	\checkmark			IFITM2	\checkmark			\checkmark	
MAL	\checkmark		\checkmark	\checkmark		IER3		\checkmark		\checkmark	
LCK	\checkmark	\checkmark		\checkmark		ID2		\checkmark		\checkmark	
KRT7			\checkmark	\checkmark	\checkmark	HPGD	\checkmark		\checkmark		
HLA-DQBI	✓			√	\checkmark	HLA-DPA I	\checkmark			\checkmark	
IL32	~	√		~		HLA-B		\checkmark		\checkmark	
ILI5	~	v		~		GZMK	\checkmark	\checkmark			
	~	\checkmark	/	~		GZMB	\checkmark	√			
	*		v	•		GZMA	~	\checkmark	/		
	v	v		1	1	GSN	~	/	✓		
	1	1		·	•	GNPDAT	*	•			
WARS	√	√ -				GINLT	v	v	./		
TSC22D1		\checkmark		\checkmark		GLS		* -	v		
TRO	\checkmark			\checkmark		GASI	•	•		1	1
TRIM22		\checkmark		\checkmark		GABARAPII			\checkmark	, ,	•
TRD@	\checkmark				\checkmark	FYN	\checkmark	\checkmark			
TRBCI	\checkmark	\checkmark				FYB		\checkmark	\checkmark		
TRATI	\checkmark	\checkmark				FLT3LG	\checkmark	\checkmark			
TPM2		\checkmark		\checkmark		FGL2	\checkmark			\checkmark	
TPMI		\checkmark		\checkmark		FER IL3				\checkmark	\checkmark
TOP2A			\checkmark		\checkmark	FAS	\checkmark			\checkmark	
TNFSF10		\checkmark		\checkmark		EPS8		\checkmark		\checkmark	
TNFAIP2	~			~		EPHX2	\checkmark		\checkmark		
TMEM158	~			~		EPHA3				\checkmark	\checkmark
	v			*		EPB41L3	√			\checkmark	
			v	•		ENPP2	\checkmark	√	,		
	* -	v			1	ENG		√	\checkmark	,	
STAT5R	×	1			•	DUSP6		v		~	
STAM	√	·			\checkmark	DUSP4	/	~		~	
SOLE	\checkmark		\checkmark				*			v	
SPOCK2	\checkmark	\checkmark					v	1		1	v
SORBS2				\checkmark	\checkmark	CYP2R4		* ✓	1	v	
SH2DIA	\checkmark	\checkmark				CYP27A1		√	•	1	
SEMA3C			\checkmark	\checkmark		CTSW	1	√ √		·	
rtni	\checkmark	\checkmark				CTSB	·	✓		\checkmark	
RGN			\checkmark	\checkmark		COL 4A3			\checkmark	✓	
RAB25			\checkmark	\checkmark		CLEC2B		\checkmark		\checkmark	
QPRT	√		\checkmark			CG018		\checkmark		\checkmark	
PTPRF	~	\checkmark			,	CD8B	\checkmark	\checkmark			
PTPRD	~				\checkmark	CD8A	\checkmark	\checkmark			
PTPN3	~			~		CD7	\checkmark	\checkmark			
PKSS23	v			*		CD6	\checkmark	\checkmark			
	1	1	v	v		CD5	\checkmark	\checkmark			
	×	v			1	CD3G	\checkmark	\checkmark			
ΡΙΡΔΚΊΟΔ	×				· ✓	CD3E	✓	✓			
PFL O	√	\checkmark				CD3D	~	~			
PEGIO			\checkmark	\checkmark		CD28	~	v			
PARD3	\checkmark				\checkmark	CD247	~	v			
NP			\checkmark	\checkmark		CD2	*	v			
NELL2	\checkmark			\checkmark			v	•			
MYCN	\checkmark		\checkmark					* -		v	1
MAST4	\checkmark			\checkmark		CCIS	1	, ,			•
MAP3K8		\checkmark			\checkmark		· •	v √			
MAFB	\checkmark	\checkmark				CAMK4	✓		\checkmark		
MAF	\checkmark	\checkmark				BCLIIB	\checkmark	\checkmark			
LTBPI		\checkmark		\checkmark		ATE3		\checkmark		\checkmark	
LEFI	~	√				ARL4C	\checkmark	\checkmark			
	v	v				AQP3	\checkmark			\checkmark	
KLKBI	✓	\checkmark		/	/	ANXAI	\checkmark			\checkmark	
			./	*	v	ANK3	\checkmark				\checkmark
		./	v	√		ANGPTI	\checkmark			\checkmark	
		* ✓		v		AKRIC2	\checkmark				\checkmark
	* -	*		\checkmark		AIFI	\checkmark		\checkmark		
II 7R	~	\checkmark		•		Genes word com	nared over the	different moto and	lyses v cha	ws that this a	one is
IL2RB	~	\checkmark				represented	ipared over the	amerent meta-dild	1/303. 7 3110	The char chils ge	
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In conclusion, the data presented reveal many novel findings regarding putative pathways within which GATA3 might act and support the hypothesis that GATA3 acts in a tissue-specific manner, while retaining its common function in differentiation and development.

Additional material on web

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References

- Mehra R, Varambally S, Ding L, Shen R, Sabel MS, Ghosh D, *et al.* Identification of GATA3 as a breast cancer prognostic marker by global gene expression meta-analysis. Cancer Res 2005;65:11259-64.
- Eeckhoute J, Keeton EK, Lupien M, Krum SA, Carroll JS, Brown M. Positive cross-regulatory loop ties GATA-3 to estrogen receptor alpha expression in breast cancer. Cancer Res 2007;67:6477-83.
- Engelsen IB, Stefansson IM, Akslen LA, Salvesen HB. GATA3 expression in estrogen receptor alpha-negative endometrial carcinomas identifies aggressive tumors with high proliferation and poor patient survival. Am J Obstet Gynecol 2008.
- Wilson BJ, Giguere V. Meta-analysis of human cancer microarrays reveals GATA3 is integral to the estrogen receptor alpha pathway. Mol Cancer 2008;7:49.
- Wilson BJ, Giguere V. Identification of novel pathway partners of p68 and p72 RNA helicases through Oncomine meta-analysis. BMC Genomics 2007;8:419.
- 6. Wilson BJ. Meta-analysis of SUMO1. BMC Res Notes 2008;1:60.
- Minegishi N, Morita S, Minegishi M, Tsuchiya S, Konno T, Hayashi N, *et al.* Expression of GATA transcription factors in myelogenous and lymphoblastic leukemia cells. Int J Hematol 1997;65:239-49.
- Chen ML, Kuo CL. A conserved sequence block in the murine and human T cell receptor Jalpha loci interacts with developmentally regulated nucleoprotein complexes in vitro and associates with GATA-3 and octamer-binding factors *in virvo*. Eur J Immunol 2001;31:1696-705.
- Marine J, Winoto A. The human enhancer-binding protein Gata3 binds to several T-cell receptor regulatory elements. Proc Natl Acad Sci U S A 1991;88:7284-8.
- Joulin V, Bories D, Eleouet JF, Labastie MC, Chretien S, Mattei MG, et al. A T-cell specific TCR delta DNA binding protein is a member of the human GATA family. Embo J 1991;10:1809-16.
- 11. Bots M, Medema JP. Granzymes at a glance. J Cell Sci 2006;119:5011-4.
- 12. Zhu J, Cote-Sierra J, Guo L, Paul WE. Stat5 activation plays a critical role in Th2 differentiation. Immunity 2003;19:739-48.
- Pandolfi PP, Roth ME, Karis A, Leonard MW, Dzierzak E, Grosveld FG, *et al.* Targeted disruption of the GATA3 gene causes severe abnormalities in the nervous system and in fetal liver haematopoiesis. Nat Genet 1995;11:40-4.
- Nardelli J, Thiesson D, Fujiwara Y, Tsai FY, Orkin SH. Expression and genetic interaction of transcription factors GATA-2 and GATA-3 during development of the mouse central nervous system. Dev Biol 1999;210:305-21.
- Pata I, Studer M, van Doorninek JH, Briscoe J, Kuuse S, Engel JD, et al. The transcription factor GATA3 is a downstream effector of Hoxb1 specification in rhombomere 4. Development 1999;126:5523-31.
- Zhao GY, Li ZY, Zou HL, Hu ZL, Song NN, Zheng MH, et al. Expression of the transcription factor GATA3 in the postnatal mouse central nervous system. Neurosci Res 2008;61:420-8.
- 17. Mukhopadhyay M, Pelka P, DeSousa D, Kablar B, Schindler A, Rudnicki

http://www.carcinogenesis.com/content/7/1/6

MA, *et al.* Cloning, genomic organization and expression pattern of a novel Drosophila gene, the disco-interacting protein 2 (dip2), and its murine homolog. Gene 2002;293:59-65.

- Akhmametyeva EM, Mihaylova MM, Luo H, Kharzai S, Welling DB, Chang LS. Regulation of the neurofibromatosis 2 gene promoter expression during embryonic development. Dev Dyn 2006;235:2771-85.
- Uppal S, Coatesworth AP. Neurofibromatosis type 2. Int J Clin Pract 2003;57:698-703.
- Van Esch H, Groenen P, Nesbit MA, Schuffenhauer S, Lichtner P, Vanderlinden G, *et al.* GATA3 haplo-insufficiency causes human HDR syndrome. Nature 2000;406:419-22.
- Muroya K, Hasegawa T, Ito Y, Nagai T, Isotani H, Iwata Y, *et al.* GATA3 abnormalities and the phenotypic spectrum of HDR syndrome. J Med Genet 2001;38:374-80.
- 22. Nesbit MA, Bowl MR, Harding B, Ali A, Ayala A, Crowe C, *et al.* Characterization of GATA3 mutations in the hypoparathyroidism, deafness and renal dysplasia (HDR) syndrome. J Biol Chem 2004;279:22624-34.
- Labastie MC, Catala M, Gregoire JM, Peault B. The GATA-3 gene is expressed during human kidney embryogenesis. Kidney Int 1995;47:1597-603.
- George KM, Leonard MW, Roth ME, Lieuw KH, Kioussis D, Grosveld F, *et al.* Embryonic expression and cloning of the murine GATA-3 gene. Development 1994;120:2673-86.
- Grote D, Souabni A, Busslinger M, Bouchard M. Pax 2/8-regulated Gata 3 expression is necessary for morphogenesis and guidance of the nephric duct in the developing kidney. Development 2006;133:53-61.
- Muto S. Potassium transport in the mammalian collecting duct. Physiol Rev 2001;81:85-116.
- Ji W, Foo JN, O'Roak BJ, Zhao H, Larson MG, Simon DB, *et al.* Rare independent mutations in renal salt handling genes contribute to blood pressure variation. Nat Genet 2008;40:592-9.
- Thompson M, Lapointe J, Choi YL, Ong DE, Higgins JP, Brooks JD, et al. Identification of candidate prostate cancer genes through comparative expression-profiling of seminal vesicle. Prostate 2008;68:1248-56.
- Perez-Stable CM, Pozas A, Roos BA. A role for GATA transcription factors in the androgen regulation of the prostate-specific antigen gene enhancer. Mol Cell Endocrinol 2000;167:43-53.
- Senoo M, Pinto F, Crum CP, McKeon F. p63 Is essential for the proliferative potential of stem cells in stratified epithelia. Cell 2007;129:523-36.
- Higgins JP, Kaygusuz G, Wang L, Montgomery K, Mason V, Zhu SX, *et al.* Placental S100 (S100P) and GATA3: Markers for transitional epithelium and urothelial carcinoma discovered by complementary DNA microarray. Am J Surg Pathol 2007;31:673-80.
- Aigner K, Dampier B, Descovich L, Mikula M, Sultan A, Schreiber M, et al. The transcription factor ZEB1 (deltaEF1) promotes tumour cell dedifferentiation by repressing master regulators of epithelial polarity. Oncogene 2007;26:6979-88.
- Anose BM, LaGoo L, Schwendinger J. Characterization of androgen regulation of ZEB-1 and PSA in 22RV1 prostate cancer cells. Adv Exp Med Biol 2008;617:541-6.
- Gregoire JM, Romeo PH. T-cell expression of the human GATA-3 gene is regulated by a non-lineage-specific silencer. J Biol Chem 1999;274:6567-78.
- 35. Bonkhoff H, Stein U, Welter C, Remberger K. Differential expression of the pS2 protein in the human prostate and prostate cancer: Association with premalignant changes and neuroendocrine differentiation. Hum Pathol 1995;26:824-8.
- Colombel M, Dante R, Bouvier R, Ribieras S, Pangaud C, Marechal JM, *et al.* Differential RNA expression of the pS2 gene in the human benign and malignant prostatic tissue. J Urol 1999;162:927-30.
- 37. Ernst T, Hergenhahn M, Kenzelmann M, Cohen CD, Bonrouhi M, Weninger A, *et al.* Decrease and gain of gene expression are equally discriminatory markers for prostate carcinoma: A gene expression analysis on total and microdissected prostate tissue. Am J Pathol 2002;160:2169-80.

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- Hosoi Y, Kapp LN, Murnane JP, Matsumoto Y, Enomoto A, Ono T, et al. Suppression of anchorage-independent growth by expression of the ataxia-telangiectasia group D complementing gene, ATDC. Biochem Biophys Res Commun 2006;348:728-34.
- Kouros-Mchr H, Slorach EM, Sternlicht MD, Werb Z. GATA-3 maintains the differentiation of the luminal cell fate in the mammary gland. Cell 2006;127:1041-55.
- Asselin-Labat ML, Sutherland KD, Barker H, Thomas R, Shackleton M, Forrest NC, *et al.* Gata-3 is an essential regulator of mammarygland morphogenesis and luminal-cell differentiation. Nat Cell Biol 2007;9:201-9.

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